

Package ‘CooccurrenceAffinity’

July 21, 2025

Type Package

Title Affinity in Co-Occurrence Data

Version 1.0.2

Date 2025-06-26

Description Computes a novel metric of affinity between two entities based on their co-occurrence (using binary presence/absence data). The metric and its MLE, α hat, were advanced in Mainali, Slud, et al, 2021 <[doi:10.1126/sciadv.abj9204](https://doi.org/10.1126/sciadv.abj9204)>. Various types of confidence intervals and median interval were developed in Mainali and Slud, 2022 <[doi:10.1101/2022.11.01.514801](https://doi.org/10.1101/2022.11.01.514801)>. The `finches` dataset is now bundled internally (no longer pulled via the cooccur package, which has been dropped).

License MIT + file LICENSE

Encoding UTF-8

LazyData true

Depends R (>= 4.1), BiasedUrn (>= 2.0.9)

Imports cowplot, ggplot2, plyr, reshape

URL <https://github.com/kpmainali/CooccurrenceAffinity>

BugReports <https://github.com/kpmainali/CooccurrenceAffinity/issues>

RoxygenNote 7.3.1

NeedsCompilation no

Author Kumar Mainali [aut, cre],
Eric Slud [aut]

Maintainer Kumar Mainali <kpmainali@gmail.com>

Repository CRAN

Date/Publication 2025-06-27 07:30:16 UTC

Contents

AcceptAffCI	2
AcceptAffin	3
affinity	4
affinity2by2	10
AlphInts	11
Bisect	14
Covrg	15
dataprep	16
EHypMidP	18
EHypQuInt	20
finches	21
logLikExtHyp	22
MaxX.Int	23
midP.EHyp	23
minmaxAlpha.pFNCH	24
MinX.Int	25
ML.Alpha	26
plotgg	28
Index	32

AcceptAffCI	<i>Acceptability Interval</i>
-------------	-------------------------------

Description

This function calculates the "Acceptability Interval" of Blaker for the log-odds parameter alpha in the Extended Hypergeometric distribution.

Usage

```
AcceptAffCI(x, marg, lev, CPint)
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
lev	a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)
CPint	the exact conservative ("Clopper-Pearson-type") interval CI.CP calculated in the function AlphInts()

Details

This function calculates the "Acceptability Interval" based on "Acceptability Function" computed by AcceptAffin(). This interval, developed by Blaker (2000), was proved in that paper's Theorem 1 in a more general class of estimation problems to have three essential properties: it falls within the CI.CP confidence interval; it maintains the property of being conservative, i.e., of having coverage probability under the Extended Hypergeometric (mA,mB,N, alpha) distribution at least as large as the nominal level; and it is larger when the confidence level is larger.

Value

This function returns the "Acceptability Interval" of Blaker (2000). The code is adapted from Blaker's Splus code for the case of an unknown binomial proportion.

Author(s)

Eric Slud

References

Blaker, H. (2000), "Confidence curves and improved exact confidence intervals for discrete distributions", Canadian Journal of Statistics 28, 783-798.

Examples

```
auxCP = AlphInts(30,c(50,80,120), lev=0.9)$CI.CP
AcceptAffCI(30,c(50,80,120), 0.9, auxCP)
```

```
AlphInts(30,c(50,80,120), lev=0.9)$CI.Blaker
```

AcceptAffin	<i>Calculates the "Acceptability Function" used in defining Blaker's (2000) Acceptability Interval and computing the latter in the function AcceptAffCI().</i>
-------------	--

Description

This function calculates the "Acceptability Function" of Blaker (2000, Thm.1, p.785) for the log-odds parameter alpha in the Extended Hypergeometric distribution.

Usage

```
AcceptAffin(x, marg, alph)
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
alph	a vector of (one or more) real-valued "alpha" values, where alpha is the log-odds parameter in the Extended Hypergeometric distribution

Details

This function calculates the "Acceptability Function" of Blaker (2000, Thm.1, p.785) for the log-odds parameter alpha in the Extended Hypergeometric distribution, a function from which the "Acceptability Interval" is calculated by another CooccurrenceAffinity package function AcceptAffCI().

Value

This function returns the "Acceptability Function" that is later used by another function AcceptAffCI() to compute "Acceptability Interval".

Author(s)

Eric Slud

References

Blaker, H. (2000), "Confidence curves and improved exact confidence intervals for discrete distributions", Canadian Journal of Statistics 28, 783-798.

affinity	<i>Computes alpha, probability, expected co-occurrence, median interval, various confidence intervals, other indices of affinity, etc.</i>
----------	--

Description

This is the principal function of "CooccurrenceAffinity" package that analyzes occurrence or abundance data (e.g., species by site) using other functions of this package and returns several quantities in one main dataframe and (optionally) up to 11 square matrices. This function processes data using dataprep() function and then feeds the data to analytical pipeline which includes ML.Alpha() and AlphInts(). The outputs of the function in the \$all dataframe include the following:

- alpha_mle: maximum likelihood estimate of the log-odds parameter alpha in the Extended Hypergeometric distribution with fixed margins (mA,mB) and table-total N, which is the "log-affinity" index of co-occurrence championed in a paper by Mainali et al. (2022) as an index of co-occurrence-based similarity; computed in ML.Alpha()

- `exp_cooccur`: expected co-occurrence count under the null (hypergeometric, corresponding to $\alpha=0$) distribution; computed as `ML.Alpha()$Null.Exp`
- `p_value`: the commonly reported P-value of the observed co-occurrences; computed by `AlphInts()$pval`
- `alpha_medianInt`: the interval of alpha values compatible with x as median for the Extended Hypergeometric distribution (Harkness 1965) with fixed margins and alpha; computed in `AlphInts()` as `$MedianIntrvl`
- `conf_level`: confidence level for estimating the various types of confidence intervals
- `ci_`: four types of confidence intervals (see details below)
- `jaccard`: Jaccard index
- `sorensen`: Sørensen-Dice index
- `simpson`: Simpson index

Usage

```
affinity(
  data,
  row.or.col,
  which.row.or.col = NULL,
  datatype = NULL,
  threshold = NULL,
  class0.rule = NULL,
  sigPval = NULL,
  sigdigit = NULL,
  squarematrix = NULL,
  ...
)
```

Arguments

<code>data</code>	occurrence matrix (binary or abundance) in matrix or dataframe format
<code>row.or.col</code>	specify if the pairs of rows or columns are analyzed for affinity. 'row' or 'column'.
<code>which.row.or.col</code>	a vector of name or the number of row/column if a subset of the data is intended to be analyzed; optional argument with default of all rows/columns.
<code>datatype</code>	specify if the datatype is 'abundance' or 'binary'; optional argument with default 'binary'.

threshold	cutoff for converting an abundance data to binary; needed if datatype is 'abundance'
class0.rule	'less.or.equal' or 'less'. 'less.or.equal' converts a threshold or lower values to zero and all the others to 1. 'less' converts a threshold and higher values to 1.
sigPval	acceptable rate of false positives or probability of rejecting the null hypothesis when it is true, commonly known as alpha in hypothesis testing
sigdigit	the number of decimals for rounding alpha mle, its intervals, expected cooccurrence under the null, jaccard, sorensen and simpson indices
squarematrix	a vector of quantities so that a square matrix for each of them is generated on the top of the main long matrix of all outputs. "alpha_mle", "alpha_mle_sig", "p_value", "cooccur.null", "cooccur.obs", "jaccard", "jaccard_sig", "sorensen", "sorensen_sig", "simpson", "simpson_sig", "all".
...	Additional arguments to control behavior of the function.

Details

This function calculates "alpha_mle", which is the maximum likelihood estimate of the log-odds parameter alpha within the Extended Hypergeometric distribution (Harkness 1965) based on the count x and fixed table margins (mA,mB) and total N, which is the "affinity" index of co-occurrence championed in the paper of Mainali et al. (2022) as an index of cooccurrence-based similarity.

This function calculates five intervals, three of them using EHypQuInt, one using EHypMidP, and one using AcceptAffCI. First ("alpha_medianInt") is the interval of alpha values compatible with x as median for the Extended Hypergeometric distribution (Harkness 1965) with fixed margins and alpha. Computed as `AlphInts()$MedianIntrvl`. This interval quantifies the underlying discreteness of the Extended Hypergeometric and its impact on the estimation of alpha. MedianIntrvl is an interval that will contain the MLE alpha-hat, and the mid-point of that interval is another reasonable estimator of alpha from the data.

There are four confidence intervals computed in `AlphInts()`, called `ci_cp`, `ci_blaker`, `ci_midQ`, `ci_midP`, matching name of the outputs in standalone outputs of `AlphInts()`, except the differences in capital/small letters. The boolean "bound" parameter is an option to prevent the intervals containing alpha-estimates to extend to plus or minus infinity, based on a Bayesian argument. The bound substituted for the Infinite endpoints is provably larger than the largest value the MLE can take whenever x avoids the endpoints $\max(mA+mB-N, 0)$ and $\min(mA, mB)$ of its logical range. The recommended confidence interval for alpha is `CI.Blaker` if a reliably conservative (over-large) coverage probability is desired, and `CI.midP` otherwise.

"ci_cp", computed as `AlphInts()$CI.CP` is an "exact" conservative test-based 2-sided confidence interval (analogous to the Clopper-Pearson (1934) confidence interval for unknown binomial proportion) for alpha based on data (x,mA,mB,N)

"ci_blaker", computed as `AlphInts()$CI.Blaker` is the Acceptability Confidence Interval of Blaker (2000, Theorem 1) which is a better confidence interval than the CP-type interval "CI.CP" in the sense of being contained within "CI.CP" but still probably conservative, i.e., with coverage probability always at least as large as the nominal level.

"ci_midQ", computed as `AlphInts()$CI.midQ` has the endpoints obtained as the midpoints of quantile intervals respectively to the $(1+\text{lev})/2$ and $(1-\text{lev})/2$ quantiles of the Extended Hypergeometric distribution.

"ci_midP", computed as `AlphInts()$CI.midQ`, behaves very similarly to "CI.midQ" and is defined by the midP approach analogous to the midP confidence interval for binomial proportions (Agresti 2013, p.605), and is calculated from `EHypMidP()`.

The recommended (slightly conservative) confidence interval is `CI.Blaker`, while the very similar intervals `CI.midQ` and `CI.midP` have coverage generally closer than `CI.CP` or `CI.Blaker` to the nominal level of coverage, at the cost of occasionally under-covering by as much as 0.04 or 0.05 for confidence levels 0.90 or 0.95. The comparison among intervals, and different possible goals that CIs of conservative or close-to-nominal coverage can serve, are similar to those compared by Brown et al. (2001) for interval estimation of an unknown binomial proportion.

"p_value" is the two-sided p-value for the equal-tailed test of the null hypothesis $\alpha=0$. This p-value is calculated when `pval="Blaker"` according to Blaker's (2000) "Acceptability" function; if the input parameter `pvalType` of `AlphInts()` is anything else, the p-value is calculated using the same idea as the midP confidence interval.

ADDITIONAL ARGUMENTS can be supplied from `ML.Alpha()` and `AlphInts()`.

Value

This function returns one main long dataframe (`$all`) with various outputs in columns (a list given under "details") for each of the pairs of the entities in row. This function also outputs optionally upto 11 square matrices of NxN entities.

Author(s)

Kumar Mainali

References

- Agresti, A. (2013) *Categorical Data Analysis*, 3rd edition, Wiley.
- Blaker, H. (2000), "Confidence curves and improved exact confidence intervals for discrete distributions", *Canadian Journal of Statistics* 28, 783-798.
- Brown, L., T. Cai, and A. DasGupta (2001), "Interval Estimation for a Binomial Proportion," *Statistical Science*, 16, 101–117.
- Clopper, C., and E. Pearson (1934), "The Use of Confidence or Fiducial Limits Illustrated in the Case of the Binomial," *Biometrika*, 26, 404–413.
- Fog, A. (2015), *BiasedUrn: Biased Urn Model Distributions*. R package version 1.07.
- Harkness, W. (1965), "Properties of the extended hypergeometric distribution", *Annals of Mathematical Statistics*, 36, 938-945.
- Mainali, K., Slud, E., Singer, M. and Fagan, W. (2022), "A better index for analysis of co-occurrence and similarity", *Science Advances*, to appear.

Examples

```
# when you have a binary presence absence occurrence data
# -----

data(finches)
head(finches)
```

```

# this dataset carries the occurrence records of 13 species in row in 17 islands in columns
dim(finches)

# the remainder of the script has been enclosed under \donttest{}
# to bypass the CRAN's 5 second limit on example files
# -----

# compute alpha and other quantities for island-pair affinity (beta diversity)
myout <- affinity(data = finches, row.or.col = "col")
myout

# you can simply flip the analysis to rows to compute species-pair affinity
myout <- affinity(data = finches, row.or.col = "row")
myout

# the rows of the outputs above include every single pair of the entities,
# producing many columns for various quantities.
# # can output an NxN square matrix for selected columns.
# an example is here
myout <- affinity(data = finches, row.or.col = "col", squarematrix = c("alpha_mle", "jaccard"))
# it is a list of three elements: one main dataframe and two square matrices
length(myout)
myout
head(myout)

# you can also compute all the square matrices with an "all"
myout <- affinity(data = finches, row.or.col = "col", squarematrix = c("all"))
# this one has 12 elements
length(myout)
myout

# when you want to compute for only certain pairs
myout <- affinity(data = finches, row.or.col = "col", which.row.or.col = 4:6,
                  squarematrix = c("alpha_mle"))
myout

myout <- affinity(data = finches, row.or.col = "col",
                  which.row.or.col = c("Isabella", "Espanola"), squarematrix = c("alpha_mle"))
print(myout)

#end of \donttest{}

# if you select only one column, the computation stops
## Not run:
myout <- affinity(data = finches, row.or.col = "col",
                  which.row.or.col = c("Darwin"), squarematrix = c("alpha_mle"))

```



```
## End(Not run)

# you can also add additional arguments bringing them from ML.Alpha() or AlphInts()
myout1 <- affinity(data = finches, row.or.col = "col",
                  which.row.or.col = c("Isabella", "Espanola"), lev=0.95, pvalType="Blaker")
myout1
myout2 <- affinity(data = finches, row.or.col = "col",
                  which.row.or.col = c("Isabella", "Espanola"), lev=0.90, pvalType="Blaker")
myout2
identical(myout1, myout2)
# myout1 and myout2 were generated with identical arguments except a difference in "lev",
# which gave different confidence intervals

myout3 <- affinity(data = finches, row.or.col = "col",
                  which.row.or.col = 4:6, lev=0.95, pvalType="Blaker")
myout3
myout4 <- affinity(data = finches, row.or.col = "col",
                  which.row.or.col = 4:6, lev=0.95, pvalType="midP")
myout4
myout3$all$p_value
myout4$all$p_value
# the p values are (or, can be) different

# when you have abundance data requiring conversion to binary
# -----
# abundance data is converted to binary based on a threshold supplied.
# it might be a good idea to explore dataprep() function and its examples
# first before workign on affinity() for abundance data.
matrix.data <- matrix(runif(400, 0, 10), nrow = 100, ncol = 4)
row.names(matrix.data) <- paste0("id_", 1:nrow(matrix.data))
colnames(matrix.data) <- paste0("variable_", 1:ncol(matrix.data))

# add some missing data and zero abundance
matrix.data[1,1] <- matrix.data[2,3] <- matrix.data[1,4] <- matrix.data[1,2] <- NA
matrix.data[10,4] <- 0
head(matrix.data)
# now this is an abundance data with some missing and some zero occurrences

# inspecting how the abundance is converted to binary first
dataprep(data = matrix.data, row.or.col = "col", datatype = "abundance",
         threshold = 5, class0.rule = "less")
myout10 <- affinity(data = matrix.data, row.or.col = "col",
                  datatype = "abundance", threshold = 5, class0.rule = "less")
myout10

# you can also feed the output of dataprep() to affinity()
myinput <- dataprep(data = matrix.data, row.or.col = "col",
                  datatype = "abundance", threshold = 5, class0.rule = "less")
myout11 <- affinity(data = myinput, row.or.col = "col", datatype = "binary")
```

```

myout11
# myout 10 and myout11 are identical
identical(myout10, myout11)

# end of \donttest{}

```

affinity2by2	<i>Maximum likelihood estimate and intervals of alpha, null expectation, p-value and traditional indices from a 2x2 table</i>
--------------	---

Description

This function uses `ML.Alpha()` and supplements to the outcome with traditional indices of Jaccard, Sorenson, and Simpson. `ML.Alpha()` calculates the maximum likelihood estimate and other quantities computed in `AlphInts()`, for the log-odds parameter alpha in the Extended Hypergeometric distribution with fixed margins (mA,mB) and table-total N, which is the "log-affinity" index of co-occurrence championed in a paper by Mainali et al. (2022) as an index of co-occurrence-based similarity.

Usage

```

affinity2by2(
  x,
  marg,
  bound = TRUE,
  scal = log(2 * marg[3]^2),
  lev = 0.95,
  pvalType = "Blaker"
)

```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
bound	a boolean parameter which when TRUE replaces the MLE of "+/-Infinity", applicable when x is respectively at the upper extreme $\min(mA, mB)$ or the lower extreme $\max(mA+mB-N, 0)$ of its possible range, by a finite value with absolute value upper-bounding the value of MLEs attainable for values of x not equal to its extremes
scal	an integer parameter (default $2 \cdot N^2$, capped at 10 within the function) that should be 2 or greater
lev	a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)

pvalType a character string telling what kind of p-value to calculate. ‘Blaker’ or “midP”. If ‘pvalType=Blaker’ (the default value), the p-value is calculated according to "Acceptability" function of Blaker (2000). If ‘pvalType=midP’, the p-value is calculated using the same idea as the midP confidence interval.

Details

See the details of `ML.Alpha()`. In addition to the output of `ML.Alpha`, this function also computes Jaccard, Sorenson and Simpson indices.

Value

This function returns maximum likelihood estimate of alpha, the interval-endpoints of alpha values for which x is a median, four confidence intervals for alpha, described in detail under documentation for `AlphInts()`, and traditional indices of Jaccard, Sorenson and Simpson. In addition there are two output list-components for the null-distribution expected co-occurrence count and the p-value for the test of the null hypothesis $\alpha=0$, calculated as in `AlphInts`.

Author(s)

Kumar Mainali and Eric Slud

References

Fog, A. (2015), BiasedUrn: Biased Urn Model Distributions. R package version 1.07.
 Harkness, W. (1965), "Properties of the extended hypergeometric distribution", *Annals of Mathematical Statistics*, 36, 938-945.
 Mainali, K., Slud, E., Singer, M. and Fagan, W. (2022), "A better index for analysis of co-occurrence and similarity", *Science Advances*, to appear.

Examples

```
ML.Alpha(x=35, c(mA=50, mB=70, N=150), lev=0.95)
affinity2by2(x=35, c(mA=50, mB=70, N=150), lev=0.95)
# ML.Alpha() is a subset of affinity2by2()
a <- ML.Alpha(x=35, c(mA=50, mB=70, N=150), lev=0.95)
b <- affinity2by2(x=35, c(mA=50, mB=70, N=150), lev=0.95)
identical(a, b[1:11])
```

AlphInts	<i>Median interval, four confidence intervals, null expectation of cooccurrence count, and p-value</i>
----------	--

Description

This function calculates (i) `MedianIntrvl`, the interval of alpha values for which the co-occurrence count is a median, (ii) four Confidence Intervals, two using `EHypQuInt()`, one using `EHypMidP()`, and one using `AcceptAffCI()`, (iii) the Expected Co-occurrence count under the Null distribution, and (iv) the p-value for the observed co-occurrence count.

Usage

```
AlphInts(x, marg, scal = log(2 * marg[3]^2), lev = 0.95, pvalType = "Blaker")
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
scal	an integer parameter (default $2 \cdot N^2$, capped at 10 within the function) that should be 2 or greater
lev	a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)
pvalType	a character string telling what kind of p-value to calculate. 'Blaker' or 'midP'. If 'pvalType=Blaker' (the default value), the p-value is calculated according to "Acceptability" function of Blaker (2000). If 'pvalType=midP', the p-value is calculated using the same idea as the midP confidence interval.

Details

This function calculates five intervals, three of them using EHypQuInt, one using EHypMidP, and one using AcceptAffCI. First ("MedianIntrvl") is the interval of alpha values compatible with x as median for the Extended Hypergeometric distribution (Harkness 1965) with fixed margins and alpha; second ("CI.CP") an "exact" conservative test-based 2-sided confidence interval (analogous to the Clopper-Pearson (1934) confidence interval for unknown binomial proportion) for alpha based on data (x,mA,mB,N); third the Acceptability Confidence Interval ("CI.Blaker") of Blaker (2000, Theorem 1) which is a better confidence interval than the CP-type interval "CI.CP" in the sense of being contained within "CI.CP" but still provably conservative, i.e., with coverage probability always at least as large as the nominal level. The fourth confidence interval ("CI.midQ") is the one given in formula (2) above of the Introduction to this documentation, with endpoints obtained as the midpoints of quantile intervals respectively to the $(1+\text{lev})/2$ and $(1-\text{lev})/2$ quantiles of the Extended Hypergeometric distribution; and the fifth ("CI.midP") which behaves very similarly to "CI.midQ" is defined by the midP approach analogous to the midP confidence interval for binomial proportions (Agresti 2013, p.605), and is calculated from EHypMidP.

The first of these intervals quantifies the underlying discreteness of the Extended Hypergeometric and its impact on the estimation of alpha. MedianIntrvl is an interval that will contain the MLE alpha-hat, and the mid-point of that interval is another reasonable estimator of alpha from the data. The recommended (slightly conservative) confidence interval is CI.Blaker, while the very similar intervals CI.midQ and CI.midP have coverage generally closer than CI.CP or CI.Blaker to the nominal level of coverage, at the cost of occasionally under-covering by as much as 0.04 or 0.05 for confidence levels 0.90 or 0.95. The comparison among intervals, and different possible goals that CIs of conservative or close-to-nominal coverage can serve, are similar to those compared by Brown et al. (2001) for interval estimation of an unknown binomial proportion.

Two other output list components are computed. First is Null.Exp, the expected co-occurrence count under the null (hypergeometric, corresponding to $\alpha=0$) distribution, and second is the two-sided p-value for the equal-tailed test of the null hypothesis $\alpha=0$. This p-value is calculated when pval="Blaker" according to Blaker's (2000) "Acceptability" function; if the input parameter pval is anything else, the p-value is calculated using the same idea as the midP confidence interval.

Value

A list of seven components: the median interval `MedianIntrvl`; the four two-sided Confidence Intervals described above, two (`CI.CP` and `CI.Blaker`) conservative and two (`CI.midQ` and `CI.midP`) with coverage probabilities generally closer to the nominal level; the null expectation `Null.Exp` of the co-occurrence count associated with $\alpha=0$; and `pval`, the two-sided p-value for the hypothesis test of $\alpha=0$, calculated by the method selected, which is the Blaker acceptability-function method if `pvalType="Blaker"` and otherwise the "midP" p-value associated with the midP confidence-interval type.

Of the four Confidence intervals produced, `CI.Blaker` is the recommended conservative interval and `CI.midP` the interval to use if coverage close to the nominal is desired.

Author(s)

Eric Slud

References

- Agresti, A. (2013) *Categorical Data Analysis*, 3rd edition, Wiley.
- Blaker, H. (2000), "Confidence curves and improved exact confidence intervals for discrete distributions", *Canadian Journal of Statistics* 28, 783-798.
- Brown, L., T. Cai, and A. DasGupta (2001), "Interval Estimation for a Binomial Proportion," *Statistical Science*, 16, 101–117.
- Clopper, C., and E. Pearson (1934), "The Use of Confidence or Fiducial Limits Illustrated in the Case of the Binomial," *Biometrika*, 26, 404–413.
- Fog, A. (2015), *BiasedUrn: Biased Urn Model Distributions*. R package version 1.07.
- Harkness, W. (1965), "Properties of the extended hypergeometric distribution", *Annals of Mathematical Statistics*, 36, 938-945.

Examples

```
unlist(AlphInts(30,c(50,80,120), lev=0.9))

AlphInts(30,c(50,80,120), lev=0.9)$CI.CP
AlphInts(30,c(50,80,120), lev=0.9)$MedianIntrvl

EHypMidP(30,c(50,80,120), 0.9)
AlphInts(30,c(50,80,120), lev=0.9)$CI.midP
# NB the third argument of AlphInts is "scal" if not named,
# so must use "lev=0.9" to define the confidence level.

EHypQuInt(30,c(50,80,120), 0.5)
AlphInts(30,c(50,80,120), lev=0.9)$MedianIntrvl

# Alpha capped warning examples
AlphInts(60,c(80,80,100), lev=0.9)
ML.Alpha(60,c(80,80,100), lev=0.9)

AlphInts(80,c(80,80,100), lev=0.9)
```

```

ML.Alpha(80,c(80,80,100), lev=0.9)

# impossible x warning examples
AlphInts(81,c(80,80,100), lev=0.9)
ML.Alpha(81,c(80,80,100), lev=0.9)

# Degenerate distribution warning example
AlphInts(80,c(80,100,100), lev=0.9)
ML.Alpha(80,c(80,100,100), lev=0.9)

```

Bisect

Bisections for finding a root of a function

Description

Find a root of a function by the method of Bisections

Usage

```
Bisect(ffnc, intrv, tol = 1e-08)
```

Arguments

ffnc	an increasing function of a single scalar argument
intrv	an interval over which the root of ffnc is sought
tol	a tolerance determining when the successive bisections of the interval within which the root will lie have become small enough to stop

Details

This function finds the root of the increasing function ffnc over the scalar interval intrv by the Method of Bisections. The function must be increasing but need not be smooth, and it must have a negative sign (value less than -tol) at the left endpoint of intrv and positive sign (value greater than tol) at the right endpoint. The method of Bisection is used in successive iterations to successively halve the width of the interval in which the root lies.

Value

This function returns a vector consisting of two numbers. The first named root is an estimate of the root x solving $\text{ffnc}(x) = 0$, valid within an error of tol. The second output vector element named fval is the value of the function ffnc at root. It should be very close to 0 unless the function happens to jump from a value less than 0 to a value greater than 0 at root.

Author(s)

Eric Slud

References

to be added

Examples

```
Bisect(function(x) x^2-1, c(0,2),1e-8)
```

Covrg	<i>Coverage Probabilities for Confidence Intervals about alpha, for fixed true alpha</i>
-------	--

Description

This function calculates the coverage probability at the true value alpha of the four types of Cpnfidence Intervals (CI.CP, CI.Blaker, CI.midQ, CI.midP) computed in AlphInts().

Usage

```
Covrg(marg, alph, scal = log(2 * marg[3]^2), lev = 0.95)
```

Arguments

- marg a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
- alph True log-odds-ratio value alpha at which coverage probabilities (under Extended Hypergeometric with parameters mA,mB,N, exp(alp)) are to be calculated
- scal an integer parameter (default 2*N^2, capped at 10 within the function) that should be 2 or greater
- lev a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)

Details

See AlphInts() documentation for details of computation of the four confidence intervals CI.CP, CI.Blaker, CI.midQ, CI.midP. The confidence intervals are calculated for each x in the allowed range from max(mA+mB-N,0) to min(mA,mB), and the probability that X=x times the indicator of alph falling in each of them is summed.

Value

A vector covPrb containing the coverage propbabilities for the four Confidence Intervals

Author(s)

Eric Slud

References

to be added

Examples

```
Covrg(c(50,70,150), 1.2, lev=0.95)
Covrg(c(50,70,150), 0, lev=0.95)
Covrg(c(50,80,120), 1.5, lev=0.9)
```

dataprep	<i>Occurrence matrix (e.g., species by site) data preparation for affinity() function</i>
----------	---

Description

This function checks the format of the data for its appropriateness, converts abundance to binary and subsets the data for the selected columns or rows. Note that the affinity can be computed between columns or between rows. In the latter case, the dataset is transposed to bring rows into the columns.

Usage

```
dataprep(
  data,
  row.or.col,
  which.row.or.col = NULL,
  datatype = NULL,
  threshold = NULL,
  class0.rule = NULL
)
```

Arguments

data	occurrence matrix (binary or abundance) in matrix or dataframe format
row.or.col	specify if the pairs of rows or columns are analyzed for affinity. 'row' or 'column'.
which.row.or.col	a vector of name or the number of row/column if a subset of the data is intended to be analyzed; optional argument with default of all rows/columns.
datatype	specify if the datatype is 'abundance' or 'binary'; optional argument with default 'binary'.
threshold	cutoff for converting an abundance data to binary; needed if datatype is 'abundance'
class0.rule	'less.or.equal' or 'less'. 'less.or.equal' converts a threshold or lower values to zero and all the others to 1. 'less' converts a threshold and higher values to 1.

Details

This function does the following:

1. checks if the supplied data is in matrix or dataframe formats which are the acceptable formats
2. if rows are selected for affinity analysis, it transposes the dataframe
3. subsets the data if specific columns or rows are selected for analysis; the selection can be made with number or name of the rows/columns
4. checks if the selected cols/rows are in numeric or integer format or not
5. checks if the selected cols/rows have data in binary 1/0 format or not; if datatype is specified as abundance, it converts it to binary format following the supplied rule

Value

A dataframe in binary 1/0 format ready to be analyzed by affinity(). Abundance data is converted to binary. A subset of the input data is returned if certain rows or columns selected. If rows are being analyzed for affinity between pairs, they are brought to columns by transposing the data.

Author(s)

Kumar Mainali

Examples

```
matrix.data <- matrix(1:40, nrow = 10, ncol = 4)

row.names(matrix.data) <- paste0("id_", 1:nrow(matrix.data))
colnames(matrix.data) <- paste0("variable_", 1:ncol(matrix.data))

# add some missing data and zero abundance
matrix.data[1,1] <- matrix.data[2,3] <- matrix.data[1,4] <- matrix.data[1,2] <- NA
matrix.data[10,4] <- 0
matrix.data
# abundance data with some missing and some zero occurrences

# some good examples
dataprep(data = matrix.data, row.or.col = "col", datatype = "abundance",
         threshold = 9, class0.rule = "less")
dataprep(data = matrix.data, row.or.col = "row", which.row.or.col = c("id_2", "id_4"),
         datatype = "abundance", threshold = 10, class0.rule = "less")
dataprep(data = matrix.data, row.or.col = "col", which.row.or.col = c("variable_1", "variable_4"),
         datatype = "abundance", threshold = 8, class0.rule = "less")
dataprep(data = matrix.data, row.or.col = "col",
         which.row.or.col = c("variable_1", "variable_3", "variable_4"),
         datatype = "abundance", threshold = 8, class0.rule = "less.or.equal")
dataprep(data = matrix.data, row.or.col = "row", datatype = "abundance",
         threshold = 10, class0.rule = "less")
dataprep(data = matrix.data, row.or.col = "col", datatype = "abundance",
         threshold = 10, class0.rule = "less")

# bad examples of specifying the rows or cols that are not in the data
```

```
## Not run:
  dataprep(data = matrix.data, row.or.col = "row",
    which.row.or.col = c("id_1", "id_4", "id_11", "id_39"), datatype = "abundance",
    threshold = 10, class0.rule = "less")
  dataprep(data = matrix.data, row.or.col = "row", which.row.or.col = c(4,7,17),
    datatype = "abundance", threshold = 10, class0.rule = "less")
  dataprep(data = matrix.data, row.or.col = "col", which.row.or.col = 2:12, datatype = "abundance",
    threshold = 10, class0.rule = "less")
  dataprep(data = matrix.data, row.or.col = "col",
    which.row.or.col = c("variable_1", "variable_9", "variable_6"), datatype = "abundance",
    threshold = 10, class0.rule = "less")

## End(Not run)

# what if you pick just one column or row
## Not run:
  dataprep(data = matrix.data, row.or.col = "row", which.row.or.col = c("id_4"),
    datatype = "abundance", threshold = 10, class0.rule = "less")

## End(Not run)

# the function fails when a required argument is missing
## Not run:
  dataprep(data = matrix.data, row.or.col = "col", which.row.or.col = c("variable_1", "variable_4"),
    datatype = "abundance", threshold = 10)
  dataprep(data = matrix.data, row.or.col = "col", which.row.or.col = c("variable_1", "variable_4"),
    datatype = "abundance", class0.rule = "less.or.equal")
  dataprep(data = matrix.data, row.or.col = "col", which.row.or.col = c("variable_1", "variable_4"),
    datatype = "abundance")

## End(Not run)

# what if you have abundance data but do not specify the datatype
## Not run:
  dataprep(data = matrix.data, row.or.col = "col", which.row.or.col = c("variable_1", "variable_4"))

## End(Not run)

# however, if it is a binary data, it's okay to not specify the datatype
# although specifying is a good practice
matrix.bindata <- dataprep(data = matrix.data, row.or.col = "col", datatype = "abundance",
  threshold = 9, class0.rule = "less")

matrix.bindata
dataprep(data = matrix.bindata, row.or.col = "col")
dataprep(data = matrix.bindata, row.or.col = "row")
```

Description

This function does the analogous calculation to that of EHypQuInt, but with the Extended Hypergeometric distribution function $F(x) = F(x, mA, mB, N, \exp(\alpha))$ replaced by $(F(x) + F(x-1))/2$.

Usage

```
EHypMidP(x, marg, lev)
```

Arguments

<code>x</code>	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
<code>marg</code>	a 3-entry integer vector (mA, mB, N) consisting of the first row and column totals and the table total for a 2x2 contingency table
<code>lev</code>	a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)

Details

This function does the analogous calculation to that of CI.CP, but with the Extended Hypergeometric distribution function $F(z, \alpha) = F(z, mA, mB, N, \exp(\alpha))$ replaced by $(F(z, \alpha) + F(z-1, \alpha))/2$.

Value

This function returns the interval of alpha values with endpoints $(F(x, \alpha) + F(x-1, \alpha))/2 = (1+lev)/2$ and $(F(x, \alpha) + F(x+1, \alpha))/2 = (1-lev)/2$.

The idea of calculating a Confidence Interval this way is analogous to the midP CI used for unknown binomial proportions (Agresti 2013, p.605).

Author(s)

Eric Slud

References

Agresti, A. (2013) Categorical Data Analysis, 3rd edition, Wiley.

Examples

```
EHypMidP(30, c(50, 80, 120), 0.9)
AlphInts(30, c(50, 80, 120), lev=0.9)$CI.midP

EHypMidP(20, c(204, 269, 2016), 0.9)
```

EHypQuInt

*Interval of alpha values for which X is a specified q'th quantile***Description**

This function outputs the largest interval of log-odds parameter values alpha for which the Extended Hypergeometric distribution function at x is $\geq q$ and the complementary distribution function $1 - F(x-)$ is $\geq 1 - q$.

Usage

```
EHypQuInt(x, marg, q, scal = log(2 * marg[3]^2))
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA + mB - N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA, mB, N) consisting of the first row and column totals and the table total for a 2x2 contingency table
q	a quantile falling strictly between 0 and 1
scal	an integer parameter (default $2 \cdot N^2$, capped at 10 within the function) that should be 2 or greater

Details

This function outputs the endpoints a1, a2 defined by

$F(x, a1) = q$ and $F(x-1, a2) = q$

where $F(z, a) = F(z, mA, mB, N, \exp(a))$ is the extended Hypergeometric distribution.

The interval of alpha values with these endpoints a1, a2 is viewed as the set of alpha values "compatible" with x being a q'th quantile for the Extended Hypergeometric.

Value

This function returns the vector (a1, a2) defined above, the endpoints of the set of alpha values for which x is a q'th quantile of the Extended Hypergeometric distribution.

Author(s)

Eric Slud

Examples

```
EHypQuInt(30, c(50, 80, 120), 0.95)
EHypQuInt(30, c(50, 80, 120), 0.05)

EHypQuInt(30, c(50, 80, 120), 0.5)
AlphInts(30, c(50, 80, 120), lev=0.9)$MedianIntrvl
```

finches

*Darwin's finches presence-absence data***Description**

A presence (1)/absence (0) matrix of seven Darwin's finch species across 13 Galápagos Islands.

Usage

```
finches
```

Format

A data frame with 7 rows (Darwin's finch species, in the row names) and 17 columns (Galápagos Islands):

Baltra presence (1) or absence (0)
Darwin presence (1) or absence (0)
Espanola presence (1) or absence (0)
Fernandina presence (1) or absence (0)
Floreana presence (1) or absence (0)
Genovesa presence (1) or absence (0)
Isabella presence (1) or absence (0)
Marchena presence (1) or absence (0)
Pinta presence (1) or absence (0)
Pinzon presence (1) or absence (0)
Rabida presence (1) or absence (0)
San.Cristobal presence (1) or absence (0)
Santa.Cruz presence (1) or absence (0)
Santa.Fe presence (1) or absence (0)
Santiago presence (1) or absence (0)
Seymour presence (1) or absence (0)
Wolf presence (1) or absence (0)

Details

Row names are the seven species: *Geospiza magnirostris*, *G. fortis*, *G. fuliginosa*, *G. difficilis*, *G. scandens*, *G. conirostris*, and *G. fuliginosa*.

Source

Sanderson J.G. (2000) Testing ecological patterns: a well-known algorithm from computer science aids the evaluation of species distributions. *American Scientist* 88, 332–339.

Griffith D.M., Veech J.A., Marsh C.J. (2016) *cooccur*: probabilistic species co-occurrence analysis in R. *Methods in Ecology and Evolution* 7, 539–543.

logLikExtHyp	<i>log of Extended Hypergeometric Likelihood at (X, mA,mB,N, alpha)</i>
--------------	---

Description

This function calculates the logarithm of the Extended Hypergeometric likelihood at specified x and alpha, with marginal totals mA, mB, N fixed.

Usage

```
logLikExtHyp(x, marg, alpha)
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
alpha	a real number, the log odds ratio or affinity parameter for the 2x2 contingency table

Details

This is simply the logarithm of the Extended Hypergeometric (Harkness 1965) or Fisher noncentral Hypergeometric, as calculated by the R package BiasedUrn. The formula is $\log(\text{pFNCHypergeo}(x, mA, N - mA, mB, \exp(\alpha)))$

Value

scalar loglikelihood value

Author(s)

Eric Slud

References

Fog, A. (2015), BiasedUrn: Biased Urn Model Distributions. R package version 1.07.
Harkness, W. (1965), "Properties of the extended hypergeometric distribution", Annals of Mathematical Statistics, 36, 938-945.

Examples

```
require(BiasedUrn)
c(logLikExtHyp(30, c(50, 80, 120), 1), log(dFNCHypergeo(30, 50, 70, 80, exp(1))))
```

MaxX.Int	<i>MaxX.Int computation</i>
----------	-----------------------------

Description

Helper function

Usage

```
MaxX.Int(marg, scal = log(2 * marg[3]^2), lev = 0.95)
```

Arguments

marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
scal	an integer parameter (default $2 \cdot N^2$, capped at 10 within the function) that should be 2 or greater
lev	a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)

Details

This is a helper function.

Value

helper function

Author(s)

Eric Slud

midP.EHyp	<i>midP.EHyp computation</i>
-----------	------------------------------

Description

Helper function

Usage

```
midP.EHyp(alp)
```

Arguments

alp	"alpha" parameter, the log-odds parameter in the Extended Hypergeometric distribution
-----	---

Details

This is a helper function.

param x integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$

Value

helper function for midP CI computation with EHypMidP

Author(s)

Eric Slud

minmaxAlpha.pFNCH	<i>integer-endpoint of range for which BiasedUrn::pFNCHHypergeo() works without error</i>
-------------------	---

Description

This function calculates an integer-endpoint of range for which BiasedUrn::pFNCHHypergeo() works without error.

Usage

```
minmaxAlpha.pFNCH(x, marg)
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table

Details

Without this function, BiasedUrn::pFNCHHypergeo() returns inconsistency message for extreme examples like: AlphInts(20,c(204,269,2016), lev=0.9, scal=10). This problem is solved within our package by restricting the range of allowed alpha to the computed (alphamin, alphamax) range.

Value

minimum and maximum of Alpha

Author(s)

Eric Slud

References

- Fog, A. (2015), BiasedUrn: Biased Urn Model Distributions. R package version 1.07.
- Harkness, W. (1965), “Properties of the extended hypergeometric distribution“, Annals of Mathematical Statistics, 36, 938-945.

Examples

```
minmaxAlpha.pFNCH(10,c(100,200,300))
minmaxAlpha.pFNCH(20,c(204,269,2016))
minmaxAlpha.pFNCH(20,c(204,269,20160))
```

MinX.Int

MinX.Int computation

Description

Helper function

Usage

```
MinX.Int(marg, scal = log(2 * marg[3]^2), lev = 0.95)
```

Arguments

- | | |
|------|--|
| marg | a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table |
| scal | an integer parameter (default $2 \cdot N^2$, capped at 10 within the function) that should be 2 or greater |
| lev | a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95) |

Details

This is a helper function.

Value

helper function

Author(s)

Eric Slud

ML.Alpha	<i>Maximum likelihood estimate and intervals of alpha, null expectation and p-value of a 2x2 table</i>
----------	--

Description

This function calculates the maximum likelihood estimate and other quantities computed in `AlphInts()`, for the log-odds parameter alpha in the Extended Hypergeometric distribution with fixed margins (mA,mB) and table-total N, which is the "log-affinity" index of co-occurrence championed in a paper by Mainali et al. (2022) as an index of co-occurrence-based similarity.

Usage

```
ML.Alpha(
  x,
  marg,
  bound = TRUE,
  scal = log(2 * marg[3]^2),
  lev = 0.95,
  pvalType = "Blaker"
)
```

Arguments

x	integer co-occurrence count that should properly fall within the closed interval $[\max(0, mA+mB-N), \min(mA, mB)]$
marg	a 3-entry integer vector (mA,mB,N) consisting of the first row and column totals and the table total for a 2x2 contingency table
bound	a boolean parameter which when TRUE replaces the MLE of "+/-Infinity", applicable when x is respectively at the upper extreme $\min(mA, mB)$ or the lower extreme $\max(mA+mB-N, 0)$ of its possible range, by a finite value with absolute value upper-bounding the value of MLEs attainable for values of x not equal to its extremes
scal	an integer parameter (default $2 \cdot N^2$, capped at 10 within the function) that should be 2 or greater
lev	a confidence level, generally somewhere from 0.8 to 0.95 (default 0.95)
pvalType	a character string telling what kind of p-value to calculate. 'Blaker' or "midP". If 'pvalType=Blaker' (the default value), the p-value is calculated according to "Acceptability" function of Blaker (2000). If 'pvalType=midP', the p-value is calculated using the same idea as the midP confidence interval.

Details

This function calculates the maximum likelihood estimate of the log-odds parameter alpha within the Extended Hypergeometric distribution (Harkness 1965) based on the count x and fixed table margins (mA,mB) and total N, which is the "affinity" index of co-occurrence championed in the

paper of Mainali et al. (2022) as an index of cooccurrence-based similarity, along with the intervals computed in `AlphInts`, called `CI.CP`, `CI.Balke`, `CI.midQ` and `CI.midP`. The boolean "bound" parameter is an option to prevent the intervals containing alpha-estimates to extend to plus or minus infinity, based on a Bayesian argument. The bound substituted for the Infinite endpoints is provably larger than the largest value the MLE can take whenever x avoids the endpoints $\max(mA+mB-N, 0)$ and $\min(mA, mB)$ of its logical range. The recommended confidence interval for alpha is `CI.Balke` if a reliably conservative (over-large) coverage probability is desired, and `CI.midP` otherwise.

Value

This function returns maximum likelihood estimate of alpha, the interval-endpoints of alpha values for which x is a median, and four confidence intervals for alpha, described in detail under documentation for `AlphInts()`. In addition there are two output list-components for the null-distribution expected co-occurrence count and the p-value for the test of the null hypothesis $\alpha=0$, calculated as in `AlphInts`.

Author(s)

Eric Slud

References

- Fog, A. (2015), `BiasedUrn`: Biased Urn Model Distributions. R package version 1.07.
- Harkness, W. (1965), "Properties of the extended hypergeometric distribution", *Annals of Mathematical Statistics*, 36, 938-945.
- Mainali, K., Slud, E., Singer, M. and Fagan, W. (2022), "A better index for analysis of co-occurrence and similarity", *Science Advances*, to appear.

Examples

```
unlist(ML.Alpha(30,c(50,80,120), lev=0.9))
AlphInts(30,c(50,80,120), lev=0.9)

AlphInts(61,c(80,80,100), lev=0.9)
ML.Alpha(61,c(80,80,100), lev=0.9)

# Alpha capped warning examples
AlphInts(60,c(80,80,100), lev=0.9)
ML.Alpha(60,c(80,80,100), lev=0.9)

AlphInts(80,c(80,80,100), lev=0.9)
ML.Alpha(80,c(80,80,100), lev=0.9)

# impossible x warning examples
AlphInts(81,c(80,80,100), lev=0.9)
ML.Alpha(81,c(80,80,100), lev=0.9)

# Degenerate distribution warning example
AlphInts(80,c(80,100,100), lev=0.9)
ML.Alpha(80,c(80,100,100), lev=0.9)
```

plotgg

*Heatmap plot of affinity() output***Description**

This function works on the output of `affinity()` and uses `ggplot2::ggplot()` to plot a heatmap plot for the numeric columns of `$all` dataframe except the interval columns (median interval and confidence intervals) and confidence level (which is a constant for all pairs in one run of the code)

Usage

```
plotgg(
  data,
  variable,
  legendlimit,
  col = NULL,
  show.value = NULL,
  value.digit = NULL,
  text.size = NULL,
  plot.margin = NULL,
  ...
)
```

Arguments

<code>data</code>	the output of <code>affinity()</code>
<code>variable</code>	a column name in <code>\$all</code> dataframe in <code>affinity()</code> output; should be a quantitative column; can be one of the following: "entity_1_count_mA", "entity_2_count_mB", "obs_cooccur_X", "sites_total_N", "p_value", "exp_cooccur", "alpha_mle", "jaccard", "sorensen", "simpson"
<code>legendlimit</code>	"datarange" or "balanced"; if "datarange", the legend spans to the range of data, if "balanced, the legend spans in equal magnitude from the center (= white by default) in both directions; note that, irrespective of the value-span in legend, the color spectrum of the plot and legend always goes from the center (=white by default) to two directions in equal magnitude. See details for more information.
<code>col</code>	a set of three colors <code>c("#87beff", "white", "#fd6a6c")</code> by default to represent low, mid and high values in the plot; these colors are applied with <code>ggplot::scale_fill_gradient2()</code>
<code>show.value</code>	a boolean to show ("TRUE") or hide ("FALSE") values in the plot; "TRUE" by default if ≤ 20 entities to compare, otherwise "FALSE" by default
<code>value.digit</code>	the number of digits in values if they are printed; default 2
<code>text.size</code>	the size of values if they are printed; default 2.5
<code>plot.margin</code>	same as <code>ggplot</code> 's <code>plot.margin</code> which includes top, right, bottom and left margins as <code>"margin(1,1,5,2, "cm")"</code>
<code>...</code>	Additional arguments to control behavior of the function.

Details

This function is really a ggplot behind the scene where I have taken care of the default value of many arguments for generating a useful plot. It generates a plot for the lower triangle of an $N \times N$ square matrix where both row and columns carry the same set of entities, such that all pairwise analyses are shown in the plot (upper triangle is the mirror image of the lower triangle and diagonals are the relation to the self which are excluded).

The plots can be requested using the column names of \$all of the main output of affinity(). The function can include additional arguments either inside plot.gg() or by appending to it with a "+" that is characteristic of ggplot().

"legendlimit" centers the legend color to white by default at null expectation in the case of alpha_mle, and negative and positive values stretch between pastel blue and pastel red colors, respectively such that the color spectrum is applied NOT to the range of data but to the same extent of values on both sides of zero, which is $\max(\text{abs}(\text{valrange}))$ and $-(\max(\text{abs}(\text{valrange})))$. For example, if alpha_mle ranges between -1.25 to 2.0, then the color spectrum always ranges between -2.0 and 2.0 but the legend can be printed to span between -1.25 and 2.0 with "dataframe" and -2.0 and 2.0 with "balanced".

For "entity_1_count_mA", "entity_2_count_mB", and "sites_total_N", there is no natural midpoint. So, "balanced" and "datarange" both use the natural behavior of ggplot in creating the color spectrum that spans between the extremes of the data.

For "obs_cooccur_X", and "exp_cooccur" also, there is no natural midpoint. To make the two plots of observed and expected cooccurrence counts comparable visually, one color scale has been applied in these two plots such that the spectrum ranges between the extremes of "obs_cooccur_X", and "exp_cooccur" collectively.

Value

This function returns a heatmap plot generated with ggplot() behind the scene.

Author(s)

Kumar Mainali

Examples

```
data(finches)
head(finches)

library(ggplot2)

# the remainder of the script has been enclosed under \donttest{}
# to bypass the CRAN's 5 second limit on example files
# -----

# plotting various variables
# -----
# compute alpha and other quantities for island-pair affinity (beta diversity)
```

```

# the square matrices are not used for plotting
myout <- affinity(data = finches, row.or.col = "col")
# myout

plotgg(data = myout, variable = "alpha_mle", legendlimit = "datarange")
# in the example above, null expectation of the alpha_mle (=0) has white color,
# and negative and positive values stretch between "#87beff" and "#fd6a6c", respectively
# so that the color spectrum is applied NOT to the range of data
# but to the same extent of values
# on both sides of zero, which is max(abs(valrange)) and -(max(abs(valrange))).
# however, the legend can be printed to show the extent of data with "datarange"
# or the entire spectrum where the color is applied with "balanced".
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced")
# notice that the two plots above are identical but the legend has
# different range with the same color scale.

plotgg(data = myout, variable = "sorensen", legendlimit = "balanced")
plotgg(data = myout, variable = "jaccard", legendlimit = "balanced")

# in the case of observed and expected cooccurrences, one color scale is applied for both plots
# so that the shades of color across plots can be visually compared
plotgg(data = myout, variable = "exp_cooccur", legendlimit = "datarange")
plotgg(data = myout, variable = "exp_cooccur", legendlimit = "balanced")
plotgg(data = myout, variable = "obs_cooccur_X", legendlimit = "balanced")

plotgg(data = myout, variable = "entity_1_count_mA", legendlimit = "datarange")
plotgg(data = myout, variable = "entity_2_count_mB", legendlimit = "datarange")
plotgg(data = myout, variable = "total_N", legendlimit = "datarange")
# for "entity_1_count_mA", "entity_2_count_mB", "sites_total_N",
# if legendlimit is set to "balanced", it will be changed to "datarange"
plotgg(data = myout, variable = "entity_2_count_mB", legendlimit = "balanced")

# change color of the plot
# -----
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced")
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced",
       col = c('#99cc33', 'white', '#ff9933'))

plotgg(data = myout, variable = "obs_cooccur_X", legendlimit = "balanced")
plotgg(data = myout, variable = "obs_cooccur_X", legendlimit = "balanced",
       col = c('#99cc33', 'white', '#ff9933'))

# change the characteristics of text printed in the plot
# -----
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced")

# change the number of digits; the default is 2
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced", value.digit = 3)

# make the fonts bigger; the default is 2.5

```

```

plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced", text.size = 3.5)

# hide values from the plot
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced", show.value = FALSE)

# increase or decrease margin
# -----

myout <- affinity(data = finches, row.or.col = "row")
# myout

plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced")
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced",
        plot.margin = ggplot2::margin(1,1,5,2, "cm"))

# change angle of x-axis tick label; the default is 35 degrees
# -----
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced")
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced") +
  ggplot2::theme(axis.text.x = element_text(angle = 45))

# to change to 90 degrees, adjust vjust
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced") +
  ggplot2::theme(axis.text.x = element_text(angle = 90))
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced") +
  ggplot2::theme(axis.text.x = element_text(angle = 90, vjust = 0.5))

# additional elements in the plot
# -----
# because it is ggplot output, you can use the arguments of ggplot() to make changes

# add plot title and change legend title
plotgg(data = myout, variable = "alpha_mle", legendlimit = "balanced") +
  ggplot2::theme(axis.text.x = element_text(angle = 90, vjust = 0.5)) +
  ggplot2::ggtitle("Affinity of island pairs measured with Alpha MLE") +
  ggplot2::labs(fill = 'Alpha MLE')

#end of \donttest{}

```

Index

* **datasets**

finches, [21](#)

AcceptAffCI, [2](#)

AcceptAffin, [3](#)

affinity, [4](#)

affinity2by2, [10](#)

AlphInts, [11](#)

Bisect, [14](#)

Covrg, [15](#)

dataprep, [16](#)

EHypMidP, [18](#)

EHypQuInt, [20](#)

finches, [21](#)

logLikExtHyp, [22](#)

MaxX.Int, [23](#)

midP.EHyp, [23](#)

minmaxAlpha.pFNCH, [24](#)

MinX.Int, [25](#)

ML.Alpha, [26](#)

plotgg, [28](#)